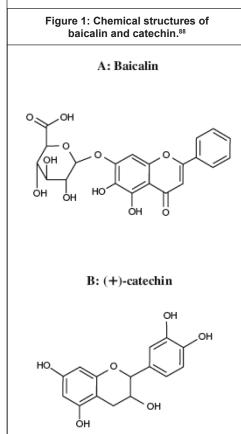
Flavocoxid: A Medicinal Extract of Scutellaria Bicalensis and Acacia Catechu Acts As a Dual Inhibitor of Cyclooxegenase and 5-Lipoxegenase to Reduce Inflammation by David M. Brady, ND, DC, CCN, DACBN

Introduction

There is great concern in the medical community over the excessive use of nonsteroidal antiinflammatory drugs (NSAIDs), including COX-2 inhibitors, and steroidal anti-inflammatory medications, as well as narcotic pain relievers, due to potentially serious side effects and dependency. In fact, patients generally use significant



doses of these as prescription drugs on a chronic or ongoing episodic basis, thereby resulting in long-term side-effect risks, particularly among the elderly. There are emerging highly standardized and evidencebased natural agents that effectively modulate the same enzyme pathways as anti-inflammatory medications with much lower side-effect profiles. One such extract with strong supporting human outcome studies, flavocoxid, will be discussed in this article with emphasis on its safety profile and balanced action across a multitude of inflammatory pathways.

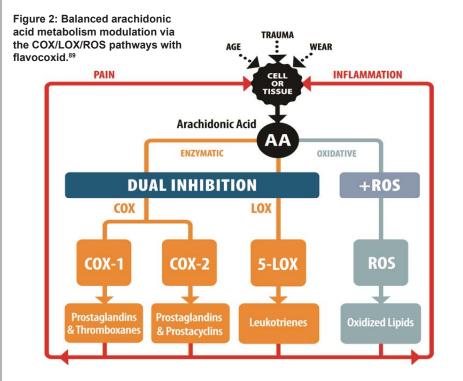
Discussion

Integrative medicine clinicians from various professional backgrounds and fields are encountering an evergrowing population of patients/ clients suffering from acute and chronic pain conditions, many of these being inflammatory in nature, including sports injuries, degenerative and inflammatory arthritis, and autoimmune-related disorders. Many such patients experience significant gastrointestinal, renal, and coagulation side effects and more, and may not even be aware of them until they cause a serious medical disorder.^{1,2} Many of these same individuals would be interested in evidence-based, effective natural agents that reduce and in many cases eliminate the drug-associated sideeffects. Integrative providers need to understand the benefits and risks of standard interventions, as well as those of the available evidence-based complementary approaches.

Some important facts about commonly used natural antiinflammatory compounds^{3–84}:

- 1. Natural anti-inflammatory compounds do not act as selective cyclooxygenase (COX) inhibitors. They constitute an array of compounds that have various combination effects of inhibiting both COX-2 and COX-1, in addition to the lipoxygenase (LOX) and phospholipase A2 enzymes. It is important that they also inhibit COX-1 to some degree because this provides a mild blood-thinning effect, counteracting the bloodclotting effect of COX-2 inhibition. In this sense, they act somewhat similar to nonselective NSAIDs (such as naproxen, diclofenac, and ibuprofen), although these medications do exert some the COX-1 dominance on isoenzyme, but they also have many additional benefits, such as antioxidant activity, and do not promote GI bleeding.
- Natural anti-inflammatory agents may be a better choice for blood thinning than aspirin, which acts predominantly as a selective COX-1 inhibitor. Aspirin binds to the *continued on page 76* ➤





platelets in an irreversible manner, with serious risk of bleeding in cases of overdose, and so comes with strong GI side effects. This is not the case with natural anti-inflammatory agents.

- 3. Natural anti-inflammatory compounds prevent the expression of "inducible" COX-2 which results from oxidative stress, due to the potent antioxidant effect of many of these compounds.
- 4. Proteolytic enzymes help reduce acute and chronic inflammation in ways unrelated to COX and LOX inhibition, such as the molecular debridement of the chemotaxispromoting protein fragments and inflammatory mediators liberated from injured cells. These enzymes also have additional antithrombotic and anti-inflammatory effects.

Figure 3: Comparative anti-inflammatory and antioxidant activity of flavocoxid and various drug agents. (Compiled by Cristiana Paul, MS, and used with permission.)

Products	Ratio of COX-1/COX-2 inhibition	Inhibits 5-LOX	Antioxidant	Side Effects		
Aspirin	15:1	NO	NO	causes blood thinning, reduces GI repair		
Ibuprofen	8:1	NO	NO	increases blood pressure, dyspepsia, GI bleeding, inhibit mitochondrial function		
Flavocoxid	1:1	YES	YES ORAC=5,517	virtually no side effects observed in clinical trials		
Rofecoxib	1:250	NO	NO	increases blood clotting, risk of heart attack		
Celecoxib	1:432	NO	NO	increases blood clotting, fisk of heart attack		
Acetaminophen	No COX1/COX2 inhibition	NO	NO (depletes glutathione)	liver toxicity		

Please see Table 1 (p. 79) for a more complete understanding of the differences between the compounds referenced above.³

Flavocoxid

Flavocoxid is specially а manufactured and extensively studied natural food-based product composed of enriched plant extracts from two botanicals, Scutellaria baicalensis and Acacia catechu. Flavocoxid contains the same "active" constituents and micronutrients that can be found in many fruits, nuts, vegetables, and teas.85 Flavonoids, low molecular weight compounds and part of the larger class of compounds known as polyphenols, are ubiquitous in plants.⁸⁶ More than 9000 different flavonoids have been characterized, many of which are consumed regularly in the human diet. Their basic structure consists of a three-

Figure 4: Improvement in Western Ontario and McMasters Universities Arthritis Index

(WOMAC) with flavocoxid vs. naproxen.94

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Flavocoxid

ring nucleus with a huge number of side chain possibilities.⁸⁶ Flavocoxid has two primary active ingredients, baicalin and catechin. Baicalin is part of a larger class of flavonoids known as free-B-ring flavonoids, whereas catechin is part of a class known as flavans (Figure 1, p. 74). The plant sources for the free-B-ring, baicalin, and the flavan, catechin, have nutritional value and have been used in medicinal preparations and in foods for many years, especially in Asian countries.

The development path for flavocoxid began in January 1996 when a scientifically based Korean ingredients supplier using pharmaceutical-like screening and development practices began to isolate multiple anti-inflammatory compounds from Aloe vera and Picrorhiza kurroa. This work was then followed by the development of high throughput (HTP) extraction and fractionation methods of plant extracts in 1998. At the same time, screening of over 1200 plant extracts and tens of thousands of HTP fractions using the COX-1, COX-2, and 5-LOX enzymes began in earnest.87 Twenty-two initial plant extracts were identified having COX-1, COX-2, and 5-LOX activity. In secondary screening in cells, the company found that 14 of the initial 22 were cytotoxic in monocytes. Further animal toxicity analysis showed that only 2 did not produce any significant changes in the physiology of animals in both acute and subchronic toxicity. These plant extracts from Scutellaria baicalensis and Acacia catechu were further characterized by HTP fractionation. They further discovered that baicalin from Scutellaria baicalensis, and catechin from Acacia catechu, were the primary active ingredients in both plant fractionations having COX-1, COX-2, and 5-LOX activity.⁸⁷

Extensive laboratory and animal toxicology testing followed in 2002-2003 characterizing the safety of the combination of naturally derived compounds.88 This science showed composing that the molecules flavocoxid at certain ratios, unlike the commonly used anti-inflammatory medications, inhibited COX-1 and COX-2 enzymes to an equal level while also inhibiting the 5-LOX enzyme (Figures 2 and 3, p. 76). These experiments in enzyme assays were also correlated with similar observations in cells. Animal models of inflammation also showed that flavocoxid could manage arachidonic acid-induced arthritis, part of the basis for osteoarthritis. In addition, later proteomic and genomic studies demonstrated that flavocoxid downregulated specific cytokines involved in the initial inflammatory cascade after cell damage due to trauma, as well as COX-2 and 5-LOX production. These results suggest that flavocoxid has both protein and genomic inhibition effects for the management of inflammation. Drug interaction studies as well as mutagenesis studies have also been performed to assure safety of the extract.89

In 2002 through early 2003, human safety and efficacy testing ensued

at Georgetown University against placebo (n = 39) for flavocoxid (n = $\frac{1}{2}$ 29) at 250 mg per day.⁸⁸ No changes in blood electrolyte, serology, liver enzymes, renal markers, or general health were observed in this study. Observed adverse events were found to be comparable to placebo. Concurrently, initial efficacy testing was done at McGill University in Canada against placebo (n = 15), flavocoxid (n = 15) at 125 mg b.i.d., flavocoxid (n = 15) at 250 mg b.i.d., and celecoxib (n = 15) at 100 mg b.i.d. Significant reductions in pain and stiffness as well as improvements in mobility were observed for flavocoxid at both doses versus celecoxib.⁹⁰ These initial safety and efficacy studies were performed and results obtained almost a year before marketing of flavocoxid as a medical food began in Puerto Rico in March 2004.

In the GOAL study, a total of 1067 individuals at 41 rheumatology practices were enrolled and prescribed flavocoxid, 500 mg b.i.d., for 60 days. The Physician Global Assessment of Disease (PGAD) visual analog scale (VAS) was used as a global measure to assess signs and symptoms of osteoarthritis (OA), including joint discomfort. functional stiffness, functional mobility, and quality of life. In, addition, GI tolerability was assessed by individual questions scored on a 5-part Likert scale. Of the 1005 patients who completed all follow-up visits, physicians recorded an average improvement in VAS scores from 60.1 \pm 18.8 at baseline to

Table 1								
Anti- inflammatory classes	COX-1 inhibition	COX-2 inhibition	Reduce inducible COX-2 expression	LOX inhibition	Anti- oxidant effect	Other anti- inflammatory effects	GI side effects, bleeding	Increases clotting and blood pressure
Natural anti-inflammatory compounds	yes	yes	yes	yes	yes	proteolytic enzyme actions	low	no
NSAIDs (naproxen, Motrin)	yes	yes	no	no	no	no	yes	no
Selective COX-2 inhibitors (Vioxx, Celebrex, Bextra)	no	yes	no	no	no	no	low	yes
Aspirin	yes	very mild	no	no	no	no	strong	no

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42.5 + 21.9 at 8 weeks (p < 0.001) in 65.8% of patients, with the highest degree of improvement seen in those subjects with moderate to severe OA at baseline. An additional important finding was that of those patients who had previously paused or interrupted their use of NSAIDs due to upper GIrelated issues, approximately 90% tolerated flavocoxid and completed the study. The use of flavocoxid also resulted in a >30% reduction or cessation of the use of gastroprotective medications such as proton pump inhibitors (PPI) or histamine-2 receptor (H2s) in subjects.⁹¹

Postmarketing Reported Adverse Events

Since the release of flavocoxid in 2004, there have been 4 reported cases of acute liver toxicity and elevated liver enzymes which are suspected to have been attributed to individual reactions to flavocoxid. All subjects were women ranging in age from 57 to 68 years. All developed symptoms and signs of liver injury within 1 to 3 months of starting flavocoxid and demonstrated elevated liver function tests (LFTs) and serum bilirubin. All serum markers returned to normal within 3 to 12 weeks after flavocoxid was discontinued, and all patients recovered without experiencing acute liver failure or chronic liver injury. Causality was adjudicated as highly likely in 3 patients and as possible in 1 patient.⁹²

have also There been 7 confirmed reports of hypersensitivity pneumonitis.⁹³ These occurred sporadically, unpredictably, and without warning or apparent predisposing factors. All required cessation of flavocoxid and treatment with supplemental oxygen and parenteral corticosteroids. This is in contrast to the reduction in respiratory adverse events of an infectious nature seen in clinical trials of flavocoxid against placebo and naproxen.94,95 These types of lung events are

extremely rare, but you should be aware of them in the unlikely event that you encounter one.

All therapeutic products that have significant physiological effects may have some level of potential toxicity and adverse effects, even a medical food with GRAS status such as flavocoxid. Considering the hundreds of thousands of prescriptions filled for flavocoxid vs. the number of reported significant side effects, it maintains an impressive safety record when compared with other antiinflammatory agents. However, it is recommended that, just as you would do with other anti-inflammatory agents or medications, you monitor liver function tests on your flavocoxid patients on the schedule that you usually use for your practice. Postmarketing surveillance on flavocoxid is collected on an ongoing basis to survey adverse events of patients while on flavocoxid. Currently, over 100,000 patients have been monitored by self-reporting or by physician-reported incidences, with an adverse event rate of $\sim 0.1\%$. An additional clinical trial sponsored by an NIH grant (Phase I SBIR Grant #: 1 R41 AR051232-01) with over 70 patients for safety at the University of Alabama-Birmingham using 250 mg b.i.d. flavocoxid versus b.i.d. placebo was published in 2009.94

Genomic/Protein Mechanism of Action (MOA) Data

Flavocoxid reduced the protein expression of COX-2 and 5-LOX, but not COX-1, presumably through an antioxidant mechanism of action (MOA) in a macrophage cell model.⁹⁶ This MOA was correlated to decreased NF κ B activation and increased I κ B α expression, the cytoplasmic control factor of NF κ B. NF κ B expression is controlled in part by oxidative activation. Tumor necrosis factor- α (TNF α) gene and protein expression as well as iNOS protein expression were also decreased through the

mechanism. Consequently, same levels of PGE2, LTB4, and nitric oxide (NO) metabolites were decreased due to the corresponding protein downregulation. Flavocoxid acted as an antioxidant, decreasing malondialdehvde (a direct oxidative of product arachidonic acid) concentrations in cell culture. These MOA data were confirmed in a Duchene muscular dystrophy (DMD) animal model by Messina et al.97

The importance of this antioxidant mechanism cannot be underestimated, especially for the management of chronic discomfort that occurs in osteoarthritis. All the molecules produced from the COX-2, 5-LOX, and iNOS pathways (e.g., leukotrienes, prostaglandins, and nitric oxide) bind to and activate the pain receptors to cause nociceptive pain signals to be transmitted back to the brain. In addition, cytokines also bind these receptors, as do a myriad of reactive oxygen species (ROS). Flavocoxid, indicated for the clinical dietary management of osteoarthritis under physician supervision, works in all these pathways and as a direct antioxidant on most of the ROS.

Head-to-Head Comparative Clinical Trials vs. Naproxen

Clinical trials of flavocoxid have continued with a well-controlled, head-to-head comparative study (n = 103) of flavocoxid 500 mg b.i.d. vs. naproxen 500 mg b.i.d. for 4 weeks, which was presented at the World Congress of the International Cartilage Repair Society (ICRS) in 2007 as a conference paper, and the complete paper has been published in the journal Nutrition Research.98 In addition, further substantiation that flavocoxid does not interact with aspirin or affect bleeding times or platelet aggregation was conducted and published by Pillai et al.99 A larger, well-controlled, head-tohead comparative study (n = 220) of flavocoxid 500 mg b.i.d. vs.

naproxen 500 mg b.i.d. for 12 weeks was then published by Levy et al., showing comparative improvements in Western Ontario and McMasters Universities Arthritis Index [WOMAC] scores (Figure 4).⁹⁴ Newer work on flavocoxid has included the study of its effect on collagen-induced arthritis in animal models, and its effect in an animal model of induced pancreatitis.^{100,101}

Commercial Availability

Flavocoxid is currently available in two commercially marketed medical food products under the supervision of a qualified licensed health-care provider: Limbrel, a medical food available in 250 mg and 500 mg versions through conventional prescription pharmacy fulfillment (Primus Pharmaceuticals Inc.: Scottsdale. Arizona, US). and Arthroben, a professionally dispensed (via licensed healthcare provider practice dispensary or authorized patient-direct order from the manufacturer) medical food product also containing novel functional collagen peptides for joint health (SitoMedica, the medical foods division of Designs for Health Inc.; Suffield, Connecticut, USA).

The concept of a "medical food" is a relatively recent one, and this product category is growing rapidly. Products now regarded as medical foods were first regulated as drugs by FDA until 1972, when the agency issued an advance notice of proposed rulemaking (ANPR), which it has since withdrawn. In 1988, Congress established the legal category of medical foods in the Orphan Drug Amendment, which states that medical foods are those designed to be orally consumed, administered under the supervision of a physician, specially formulated and processed (i.e., cannot be derived at the given concentration or formulation through a change in diet alone), and intended for the specific dietary management of a disease or condition that has

distinctive nutritional requirements. That definition was subsequently incorporated into the Nutrition Labeling and Education Act of 1990 (NLEA), wherein Congress exempted medical foods from the nutrition labeling, health claim, and nutrient disclosure requirements applied to most other foods. This has created a territory somewhere between supplement and drug where products, if they contain only ingredients with GRAS (generally recognized as safe) status by FDA as supported by robust dossiers of consensus scientific support, including published direct human outcome data as well as toxicology data, and are labeled for the dietary management of a specific disease or condition that has distinctive nutritional requirements. can then be marketed with the supported medical claims. These products are expressly required to follow "good scientific principles," which broadly include being supported by well-controlled clinical and scientific studies (using the formulation in the finished product) recognized by experts in the field such as peer review in recognized medical and scientific journals. This lifts the bar for natural products well beyond what is currently required to produce and market a product in the nutritional supplement regulatory category, where the manufacturer is extremely restricted in making any disease or health-related claims for the product, even when support for the individual ingredients may exist in the literature.

Conclusion

While there are many traditionally used natural agents with antiinflammatory properties, there are not many with rigorous scientific studies proving safety and efficacy that meet the standards of classification as a medical food. Flavocoxid is such a natural material and provides the integrative clinician with a reliable, proven, low side–effect profile, natural option for the dietary management of metabolic inflammatory processes in conditions such as osteoarthritis.

Disclosures

Dr. David M. Brady is the chief medical officer of Designs for Health Inc. and its subsidiary division SitoMedica, and is also a clinical consultant to Genova Diagnostics.

Notes

- Singh G, Ramey RD, Morfeld D, et al. Gastrointestinal tract complications of nonsteroidal anti-inflammatory drug treatment in rheumatoid arthritis. A prospective observational cohort study. Atch Inter Med. 156;1530–1536; July 1996.
- Nasrallah R, Robertson SJ, et al. Celecoxib modifies glomerular basement membrane, mesangium and podocytes in OVE26 mice, but ibuprofen is more detrimental. *Clin Sci (Lond)*. 2013 Jun;124(11):685–694.
- Brady DM, Paul C. Anti-inflammatory options: Vioxx and other secretive COX-2 inhibitors v natural agents. *Nutr Perspectives*. 2009;32(2):33– 36.
- Ammon HP, Safayhi H. Mechanism of antiinflammatory actions of curcumin and boswellic acids. J Ethnopharmacol. 1993 Mar;38(2–3):113– 119.
- Safayhi H, Mack T. Boswellic acids: novel, specific, nonredox inhibitors of 5-lipoxygenase. J Pharmacol Exp Ther. 1992 Jun;261(3):1143–1146.
- Ammon HP, Mack T. Inhibition of leukotriene B4 formation in rat peritoneal neutrophils by an ethanolic extract of the gum resin exudate of Boswellia serrata. *Planta Med.* 1991 Jun; 57(3):203–207.
- Safayhi H, Sailer ER. Mechanism of 5-lipoxygenase inhibition by acetyl-11-keto-beta-boswellic acid. *Mol Pharmacol.* 1995 Jun;47(6):1212–1216.
- Huang MT, Badmaev V. Anti-tumor and anticarcinogenic activities of triterpenoid, betaboswellic acid. *Biofactors*. 2000;13(1–4): 225–30.
- Ammon HP. Boswellic acids (components of frankincense) as the active principle in treatment of chronic inflammatory diseases *Wien Med Wochenschr.* 2002;152(15–16):373–378.
- Wallace JM. Nutritional and botanical modulation of the inflammatory cascade—eicosanoids, cyclooxygenases, and lipoxygenases—as an adjunct in cancer therapy. *Integr Cancer Ther.* 2002 Mar; 1(1):7–37; discussion 37.
- Kiuchi F, Iwakami S. Inhibition of prostaglandin and leukotriene biosynthesis by gingerols and diarylheptanoids. *Chem Pharm Bull (Tokyo)*. 1992 Feb;40(2):387–391.
- 12. Koo KL, Ammit AJ. Gingerols and related analogues inhibit arachidonic acid-induced human platelet serotonin release and aggregation. *Thromb Res.* 2001 Sep 1;103(5):387–397.
- Nurtjahja-Tjendraputra E. Effective anti-platelet and COX-1 enzyme inhibitors from pungent constituents of ginger. *Thromb Res.* 2003;111(4– 5):259–265.
- 14. Surh YJ. Anti-tumor promoting potential of selected spice ingredients with antioxidative and anti-inflammatory activities: a short review. *Food Chem Toxicol*. 2002 Aug;40(8):1091–1097.

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- 15. Murakami A, Takahashi D. Zerumbone, a Southeast Asian ginger sesquiterpene, markedly suppresses free radical generation, proinflammatory protein production, and cancer cell proliferation accompanied by apoptosis: the alpha,betaunsaturated carbonyl group is a prerequisite. *Carcinogenesis.* 2002 May;23(5):795–802.
- Murakami A, Hayashi R. Suppression of dextran sodium sulfate-induced colitis in mice by zerumbone, a subtropical ginger sesquiterpene, and nimesulide: separately and in combination. *Biochem Pharmacol.* 2003 Oct 1;66(7):1253– 1261.
- Lumb AB. Effect of dried ginger on human platelet function. *Thromb Haemost*. 1994 Jan;71(1): 110– 111.
- Janssen PL, Meyboom S. Consumption of ginger (Zingiber officinale roscoe) does not affect ex vivo platelet thromboxane production in humans. *Eur J Clin Nutr.* 1996 Nov;50(11): 772–774.
- 19. Verma SK, Singh J. Effect of ginger on platelet aggregation in man. *Indian J Med Res.* 1993 Oct;98: 240–242.
- Srivastava KC. Effect of onion and ginger consumption on platelet thromboxane production in humans. *Prostaglandins Leukot Essent Fatty Acids.* 1989 Mar;35(3):183–185.
- Bierhaus A, Zhang Y. The dietary pigment curcumin reduces endothelial tissue factor gene expression by inhibiting binding of AP-1 to the DNA and activation of NF-kappa B. Thromb Haemost. 1997 Apr;77(4):772–782.
- 22. Shah BH, Nawaz Z. Inhibitory effect of curcumin, a food spice from turmeric, on platelet-activating factor- and arachidonic acid-mediated platelet aggregation through inhibition of thromboxane formation and Ca2+ signaling. *Biochem Pharmacol.* 1999 Oct 1;58(7):1167–1172.
- Srivastava KC, Bordia A. Curcumin, a major component of food spice turmeric (Curcuma longa) inhibits aggregation and alters eicosanoid metabolism in human blood platelets. *Prostaglandins Leukot Essent Fatty Acids*. 1995 Apr;52(4):223–227.
- Ammon HP, Safayhi H. Mechanism of antiinflammatory actions of curcumine and boswellic acids. *J Ethnopharmacol.* 1993 Mar;38(2–3):113–119.
- Goel A, Boland CR. Specific inhibition of cyclooxygenase-2 (COX-2) expression by dietary curcumin in HT-29 human colon cancer cells. *Cancer Lett.* 2001 Oct 30;172(2):111–118.
- Surh YJ. Anti-tumor promoting potential of selected spice ingredients with antioxidative and anti-inflammatory activities: a short review. *Food Chem Toxicol.* 2002 Aug;40(8):1091–1097
- Surh YJ, Chun KS. Molecular mechanisms underlying chemopreventive activities of antiinflammatory phytochemicals: down-regulation of COX-2 and iNOS through suppression of NFkappa B activation. *Mutat Res.* 2001 Sep 1;480– 481:243–268.
- Sharma RA, Ireson CR. Effects of dietary curcumin on glutathione S-transferase and malondialdehyde-DNA adducts in rat liver and colon mucosa: relationship with drug levels. *Clin Cancer Res.* 2001 May;7(5):1452–1458.
- Rao CV, Kawamori T. Chemoprevention of colonic aberrant crypt foci by an inducible nitric oxide synthase-selective inhibitor. *Carcinogenesis*. 1999 Apr;20(4):641–644.
- Pendurthi UR, Williams JT. Inhibition of tissue factor gene activation in cultured endothelial cells by curcumin. Suppression of activation of transcription factors Egr-1, AP-1, and NFkappa B. Arterioscler Thromb Vasc Biol. 1997 Dec;17(12):3406–3413.

- Satoskar RR, Shah SJ. Evaluation of antiinflammatory property of curcumin (diferuloyl methane) in patients with postoperative inflammation. Int J Clin Pharmacol Ther Toxicol. 1986 Dec;24(12):651–654.
- Thamlikitkul V, Bunyapraphatsara N. Randomized double blind study of Curcuma domestica Val. for dyspepsia. *Med Assoc Thai*. 1989 Nov;72(11):613–620.
- Kobayashi T, Hashimoto S. Curcumin inhibition of Dermatophagoides farinea-induced interleukin-5 (IL-5) and granulocyte macrophage-colony stimulating factor (GM-CSF) production by lymphocytes from bronchial asthmatics. *Biochem Pharmacol.* 1997 Oct 1;54(7):819–824.
- Ram A, Das M. Curcumin attenuates allergeninduced airway hyperresponsiveness in sensitized guinea pigs. *Biol Pharm Bull.* 2003 Jul;26(7):1021– 1024.
- Masuda T, Hidaka K. Chemical studies on antioxidant mechanism of curcuminoid: analysis of radical reaction products from curcumin. J Agric Food Chem. 1999 Jan;47(1):71–77.
- Kapoor S, Priyadarsini KI. Protection of radiationinduced protein damage by curcumin. *Biophys Chem.* 2001 Aug 30;92(1–2):119–126.
- Sun YM, Zhang HY. Theoretical elucidation on the antioxidant mechanism of curcumin: a DFT study. Org Lett. 2002 Aug 22;4(17):2909–2911.
- Masuda T, Toi Y. Structural identification of new curcumin dimers and their contribution to the antioxidant mechanism of curcumin. J Agric Food Chem. 2002 Apr 24;50(9):2524–2530.
- 39. Luo F, Huang R. Protective effect and mechanism of pretreatment with curcumin on infectious brain edema in rats. *Zhonghua Er Ke Za Zhi.* 2003 Dec;41(12):940–944.
- Aleksandrov PN, Speranskaia TV. Effect of rutin and esculamine on models of aseptic inflammation. [Article in Russian.] Farmakol Toksikol. 1986 Jan–Feb;49(1):84–86.
- Guardia T, Rotelli AE. Anti-inflammatory properties of plant flavonoids. Effects of rutin, quercetin and hesperidin on adjuvant arthritis in rat. *Farmaco*. 2001 Sep;56(9):683–687.
- Olszanecki R, Gebska A. Flavonoids and nitric oxide synthase. J Physiol Pharmacol. 2002 Dec;53(4 Pt 1):571–584.
- Kiho T, Usui S, et al. Tomato paste fraction inhibiting the formation of advanced glycation end-products. *Biosci Biotechnol Biochem*. 2004;68(1):200–205.
- Vadas P, Stefanski E. Potential therapeutic efficacy of inhibitors of human phospholipase A2 in septic shock. Agents Actions. 1986 Nov;19(3–4):194– 202.
- Gryglewski RJ, Korbut R. On the mechanism of antithrombotic action of flavonoids. *Biochem Pharmacol.* 1987 Feb 1;36(3):317–222.
- Swies J, Robak J. Antiaggregatory effects of flavonoids in vivo and their influence on lipoxygenase and cyclooxygenase in vitro. *Pol J Pharmacol Pharm.* 1984 Sep–Oct;36(5):455–463.
- 47. Aleksandrov PN, Speranskaia TV. Op cit.
- 48. Guardia T, Rotelli AE. Op cit.
- Formica JV, Regelson W. Review of the biology of Quercetin and related bioflavonoids. *Food Chem Toxicol.* 1995 Dec;33(12): 1061–1680.
- Stefanescu M, Matache C. Modulation of cell adhesion by tyrosine kinases and phosphatases inhibitors. *Roum Arch Microbiol Immunol.* 1997 Jan–Jun;56(1–2):3–15.
- Fitzpatrick DF, Hirschfield SL. Endotheliumdependent vasorelaxing activity of wine and other grape products. *Am J Physiol.* 1993 Aug;265(2 Pt 2):H774–H778.

- Otsuka H, Inaba M. Histochemical and functional characteristics of metachromatic cells in the nasal epithelium in allergic rhinitis: studies of nasal scrapings and their dispersed cells. J Allergy Clin Immunol. 1995 Oct;96(4):528–536.
- Pearce FL, Befus AD, Bienenstock J. Mucosal mast cells. III. Effect of quercetin and other flavonoids on antigen-induced histamine secretion from rat intestinal mast cells. J Allergy Clin Immunol. 1984 Jun;73(6): 819–823.
- Middleton E Jr, Drzewiecki G. Flavonoid inhibition of human basophil histamine release stimulated by various agents. *Biochem Pharmacol*. 1984 Nov 1;33(21):3333–3338.
- Kimata M, Shichijo M. Effects of luteolin, quercetin and baicalein on immunoglobulin E-mediated mediator release from human cultured mast cells. *Clin Exp Allergy*. 2000 Apr;30(4): 501–8.
- O'Prey J, Brown J. Effects of dietary flavonoids on major signal transduction pathways in human epithelial cells. *Biochem Pharmacol.* 2003 Dec 1;66(11):2075–2088.
- Thuillier P, Brash AR. Inhibition of peroxisome proliferator-activated receptor (PPAR)-mediated keratinocyte differentiation by lipoxygenase inhibitors. *Biochem J.* 2002 Sep 15;366(Pt 3):901– 910.
- da Silva EL, Abdalla DSP, Terao J. Inhibitory effect of flavonoids on low-density lipoprotein peroxidation catalyzed by mammalian 15-lipoxygenase. *IUBMB Life*. 2000;49:289–295.
- Miodini P, Fioravanti L. The two phyto-oestrogens genistein and quercetin exert different effects on oestrogen receptor function. *Br J Cancer.* 1999 Jun;80(8):1150–1155.
- Koga T, Meydani M. Effect of plasma metabolites of (+)-catechin and quercetin on monocyte adhesion to human aortic endothelial cells. *Am J Clin Nutr.* 2001 May;73(5):941–948.
- De Whalley CV, Rankin SM. Flavonoids inhibit the oxidative modification of low density lipoproteins by macrophages. *Biochem Pharmacol.* 1990 Jun 1;39(11):1743–1750.
- 62. Teixeira S. Bioflavonoids: proanthocyanidins and quercetin and their potential roles in treating musculoskeletal conditions. J Orthop Sports Phys Ther. 2002 Jul;32(7):357–363.
- Lo AH, Liang YC. Carnosol, an antioxidant in rosemary, suppresses inducible nitric oxide synthase through down-regulating nuclear factorkappaB in mouse macrophages. *Carcinogenesis*. 2002 Jun;23(6):983–991.
- 64. Chan MM, Ho CT. Effects of three dietary phytochemicals from tea, rosemary and turmeric on inflammation-induced nitrite production. *Cancer Lett.* 1995 Sep 4;96(1):23–29.
- Huang MT, Ho CT. Inhibition of skin tumorigenesis by rosemary and its constituents carnosol and ursolic acid. *Cancer Res.* 1994 Feb 1;54(3):701– 708.
- Wargovich MJ, Woods C. Herbals, cancer prevention and health. J Nutr. 2001 Nov;131(11 Suppl):3034S–30346S
- Singletary K, MacDonald C. Inhibition by rosemary and carnosol of 7,12-dimethylbenz[a]anthracene (DMBA)-induced rat mammary tumorigenesis and in vivo DMBA-DNA adduct formation. *Cancer Lett.* 1996 Jun 24;104(1):43–48.
- Kosaka K, Yokoi T. Carnosic acid, a component of rosemary (Rosmarinus officinalis L.), promotes synthesis of nerve growth factor in T98G human glioblastoma cells.
- Danilenko M, Wang X. Carnosic acid and promotion of monocytic differentiation of HL60-G cells initiated by other agents. *J Natl Cancer Inst.* 2001 Aug 15;93(16):1224–1233.

- Valenzuela A, Sanhueza J. Cholesterol oxidation: health hazard and the role of antioxidants in prevention. *Biol Res.* 2003;36(3–4):291–302.
- Debersac P, Vernevaut MF. Effects of a watersoluble extract of rosemary and its purified component rosmarinic acid on xenobioticmetabolizing enzymes in rat liver. *Food Chem Toxicol.* 2001 Feb;39(2):109–117.
- 72. Singletary KW, Rokusek JT. Tissue-specific enhancement of xenobiotic detoxification enzymes in mice by dietary rosemary extract. *Plant Foods Hum Nutr.* 1997;50(1):47–53.
- Singletary KW. Rosemary extract and carnosol stimulate rat liver glutathione-S-transferase and quinone reductase activities. *Cancer Lett.* 1996 Feb 27;100(1–2):139–144.
- Del Campo J, Amiot MJ. Antimicrobial effect of rosemary extracts. J Food Prot. 2000 Oct;63(10):1359–1368
- Culpitt SV, Rogers DF. Inhibition by red wine extract, resveratrol, of cytokine release by alveolar macrophages in COPD. *Thorax.* 2003 Nov;58(11):942–946.
- Cavallaro A, Ainis T. Effect of resveratrol on some activities of isolated and in whole blood human neutrophils. *Physiol Res.* 2003;52(5): 555–562.
- 77. Ignatowicz E, Baer-Dubowska W. Resveratrol, a natural chemopreventive agent against degenerative diseases. *Pol J Pharmacol*. 2001 Nov–Dec;53(6):557–569.
- Bertelli AA, Baccalini R. Resveratrol inhibits TNF alpha-induced endothelial cell activation. *Therapie*. 2001 Sep–Oct;56(5):613–616.
- Manna SK, Mukhopadhyay A. Resveratrol suppresses TNF-induced activation of nuclear transcription factors NF-kappa B, activator protein-1, and apoptosis: potential role of reactive oxygen intermediates and lipid peroxidation. J Immunol. 2000 Jun 15;164(12):6509–6519.
- Tsai SH, Lin-Shiau SY. Suppression of nitric oxide synthase and the down-regulation of the activation of NFkappaB in macrophages by resveratrol. Br J Pharmacol. 1999 Feb:126(3):673–680.
- Pace-Asciak CR, Hahn S. The red wine phenolics trans-resveratrol and quercetin block human platelet aggregation and eicosanoid synthesis: implications for protection against coronary heart disease. *Clin Chim Acta*. 1995 Mar 31;235(2):207– 219.
- 82. Sadik CD, Sies H. Inhibition of 15-lipoxygenases by flavonoids: structure-activity relations and

mode of action. *Biochem Pharmacol*. 2003 Mar 1;65(5):773-781.

- FitzGerald GA, Patrono C. The coxibs, selective inhibitors of cyclooxygenase-2. N Engl J Med. 2001 Aug 9;345(6):433–442.
- De Caterina R, Zampolli A. From asthma to atherosclerosis – 5-lipoxygenase, leukotrienes, and inflammation. N Engl J Med. 2004 Jan 1;350(1):4– 7.
- USDA national nutrient database for standard reference: micronutrient analysis [Web page]. 2003. http://ndb.nal.usda.gov.
- Middleton E Jr., Kandaswami C, Theoharides TC. The effects of plant flavonoids on mammalian cells: implications for inflammation, heart disease, and cancer. *Pharmacol Rev*.2000 Dec;52(4):673– 751.
- Jia Q. 2003. Generating and screening a natural product library for cyclooxygenase & lipoxygenase dual inhibitors. *Stud Nat Product Chem.* 29:643– 718.
- Burnett BP, Jia Q, Zhao Y, and Levy RM. A medicinal extract of Scutellaria baicalensis and Acacia catechu acts as a dual inhibitor of cyclooxygenase and 5-lipoxygenase to reduce inflammation. *J Medicinal Foods*. 2007;10:442– 451.
- Burnett BP, Bitto A, Squadrito F, Levy RM, Pillai L. 2011. Flavocoxid inhibits phospholipase A2, peroxidase moieties of the cyclooxygenases (COX), 5-lipoxygenase, modifies COX-2 gene expression and acts as an antioxidant. *Mediators Inflamm.* 2011:385780. Epub June 22. doi:10.1155/2011/385780.
- Sampalis JS, Brownell LA. A randomized, double blind, placebo and active comparator controlled pilot study of UP446, a novel dual pathway inhibitor anti-inflammatory agent of botanical origin. Nutr J. 2012 Apr 5;11:21.
- Pillai L, Burnett BP, Levy RM. GOAL: multicenter, open-label, post-marketing study of flavocoxid, a novel dual pathway inhibitor anti-inflammatory agent of botanical origin. *Curr Med Res Opin*. 2010 May;26(5):1055–1063.
- Chalasani N, Vuppalanchi R, Navarro V, et al. Acute liver injury due to flavocoxid (Limbrel), a medical food for osteoarthritis: a case series. Ann Intern Med. 2012 June 19;156(12);857–860.
- Panduranga V, Atienza J, Kumar A, et al. Hypersensitivity pneumonitis due to flavocoxid: are corticosteroids necessary? *Conn Med.* 2013 Feb;77(2);87–90.

- Morgan SL, Baggott JE, Moreland E, et al. The safety of flavocoxid, a medical food, in the dietary management of knee osteoarthritis. J Med Food. 2009 Oct;12(5);1143–1148.
- Levy RM, Khokhlov A, Kopenken S, et al. Efficacy and safety of flavocoxid, a novel therapeutic, compared with naproxen: a randomized multicenter controlled trial in subjects with osteoarthritis of the knee. Adv Ther. 2010 Oct;27(10):731–742.
- 96. Altavilla D, Bitto A, Polito F, et al. Flavocoxid, a dual inhibitor of cyclooxygenase and 5-lipoxygenase, blunts pro-inflammatory phenotype activation in endotoxin stimulated macrophages. *Brit J Pharmacol.* 2009;157:1410– 1418.
- Messina S, Bitto A, Aquennouse M, et al. Flavocoxid counteracts muscle necrosis and improves functional properties in mdx mice: a comparison study with methylprednisolone. *Exp Neurol.* 2009;Dec;220(2):349–358.
- Levy RM, Saikovsky R, Schmidt E, et al. Flavocoxid is as effective as naproxen for managing the signs and symptoms of osteoarthritis of the knee in humans: a short-term randomized, double-blind pilot study. Nutr Res. 2009;May;29(5):298–304.
- Pillai L, Levy RM, Yiman M, et al. Flavocoxid, an anti-inflammatory agent of botanical origin, does not affect coagulation or interact with anticoagulation therapies. *Adv Ther.* 2010 Jun;27(6):400–411.
- 100. Polito F, Irrera N, Bitto A, et al. Flavocoxid, a dual inhibitor of COX and 5-LOX, attenuates inflammation and bone resorption in a mouse model of collagen induced arthritis. 2011. Data on file at Primus Pharmaceuticals Inc., Scottsdale, AZ.
- 101. Polito F, Bitto A, Irrera N, et al. Flavocoxid, a dual inhibitor of cyclooxygenase-2 and 5-lipoxygenase, reduces pancreatic damage in an experimental model of acute pancreatitis. Br J Pharmacol. 2010 Nov;161(5):1002–1011.

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